

Gene editing livestock

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ISBR 2025

Gene edited livestock with positive regulatory decisions





TOWARDS IMPROVEMENT OF **RUMINANT** BREEDING
THROUGH **GENOMIC** AND EPIGENOMIC APPROACHES

RUMIGEN, coordinated by INRAE, involves 18 partners in 7
Member states plus Norway and United Kingdom



Start date: 1st June 2021



End date: 31st May 2026



EU contribution: 6.998.851 €



Project coordinator: Eric Pailhoux



Biotechnology and
Biological Sciences
Research Council



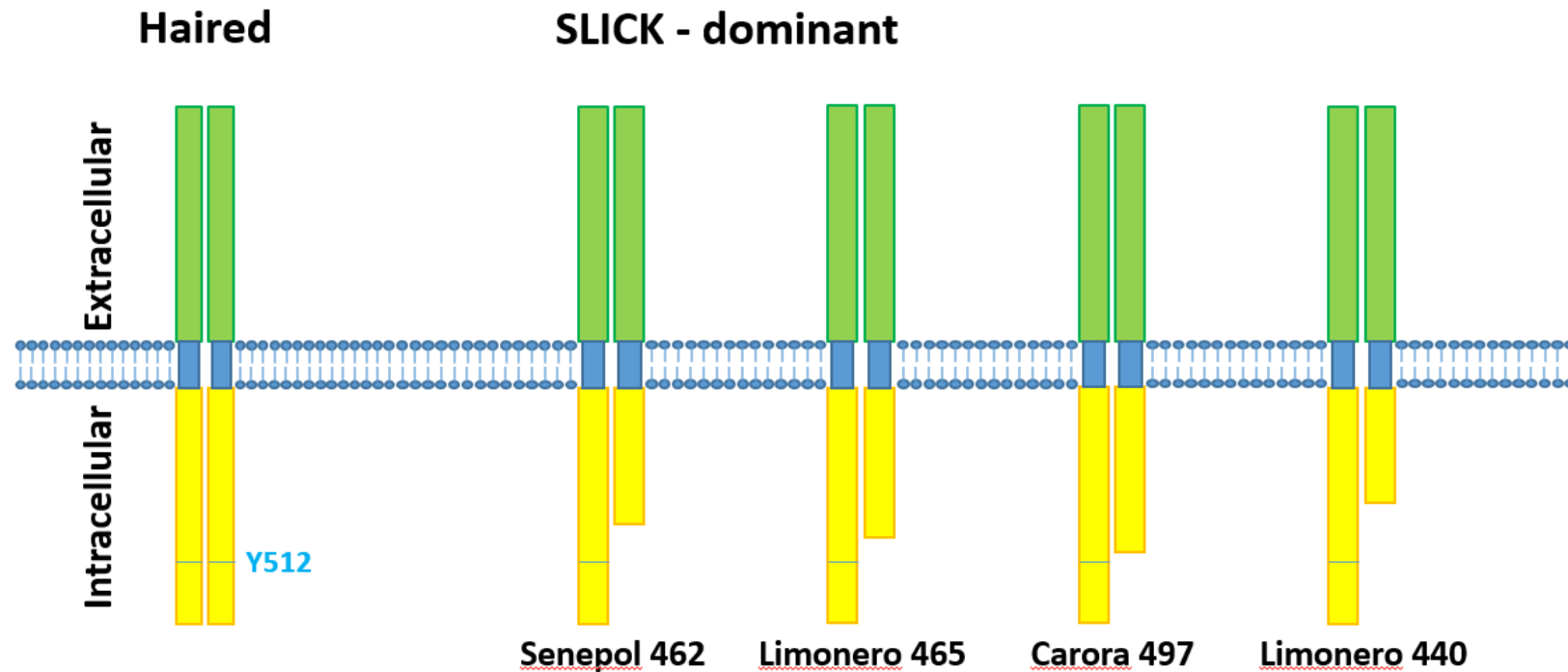
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The slick phenotype in cattle

Mutations in the prolactin receptor (PRLR) gene are associated with improved thermal tolerance in cattle



Y512 dimerization required for Jak2/STAT5 activation

(adapted from Porto-Neto et al., 2018)

Will PRLR mutation in sheep produce a slick phenotype?

Control (RUM02)



Homozygote (RUM03)



Will PRLR mutation in sheep produce a slick phenotype?

Control (RUM02)



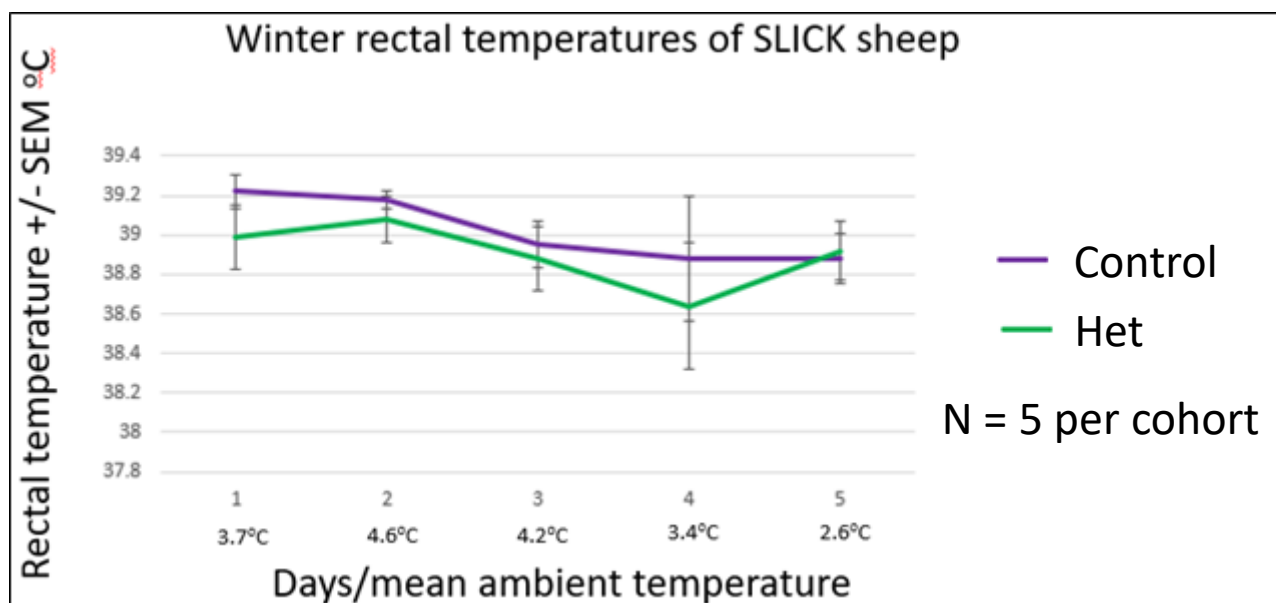
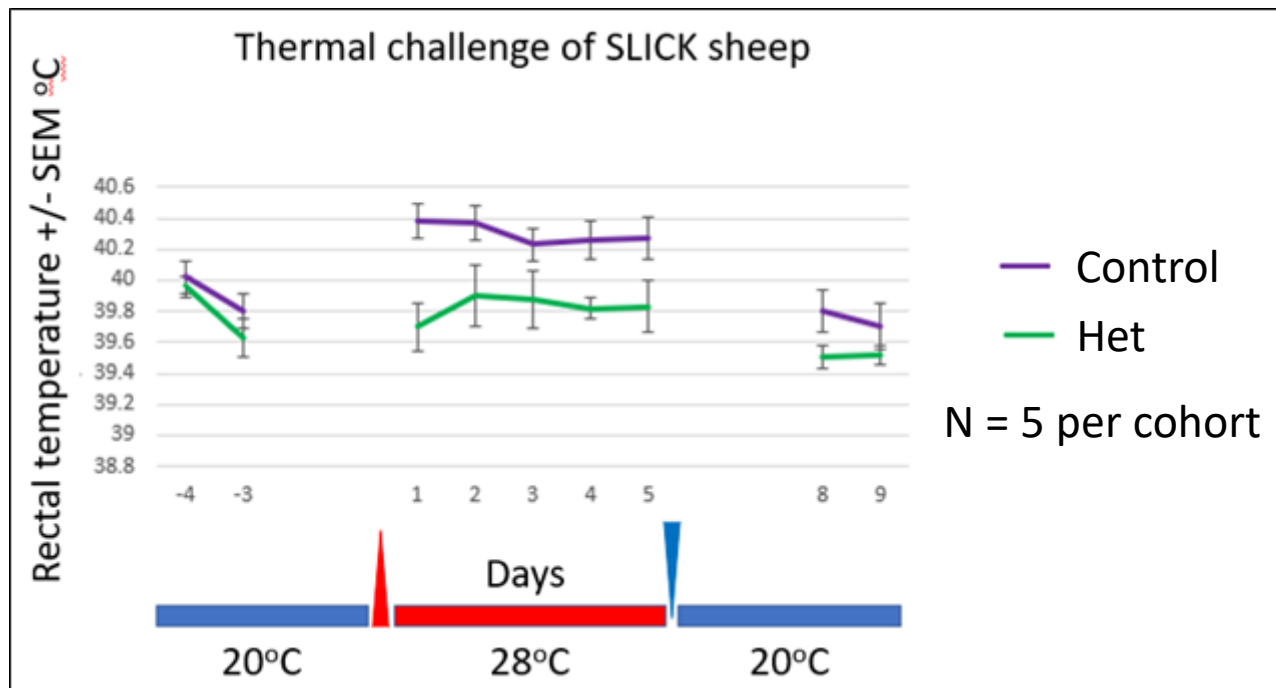
Homozygote (RUM03)



August
6 months old

SLICK sheep have improved thermal tolerance

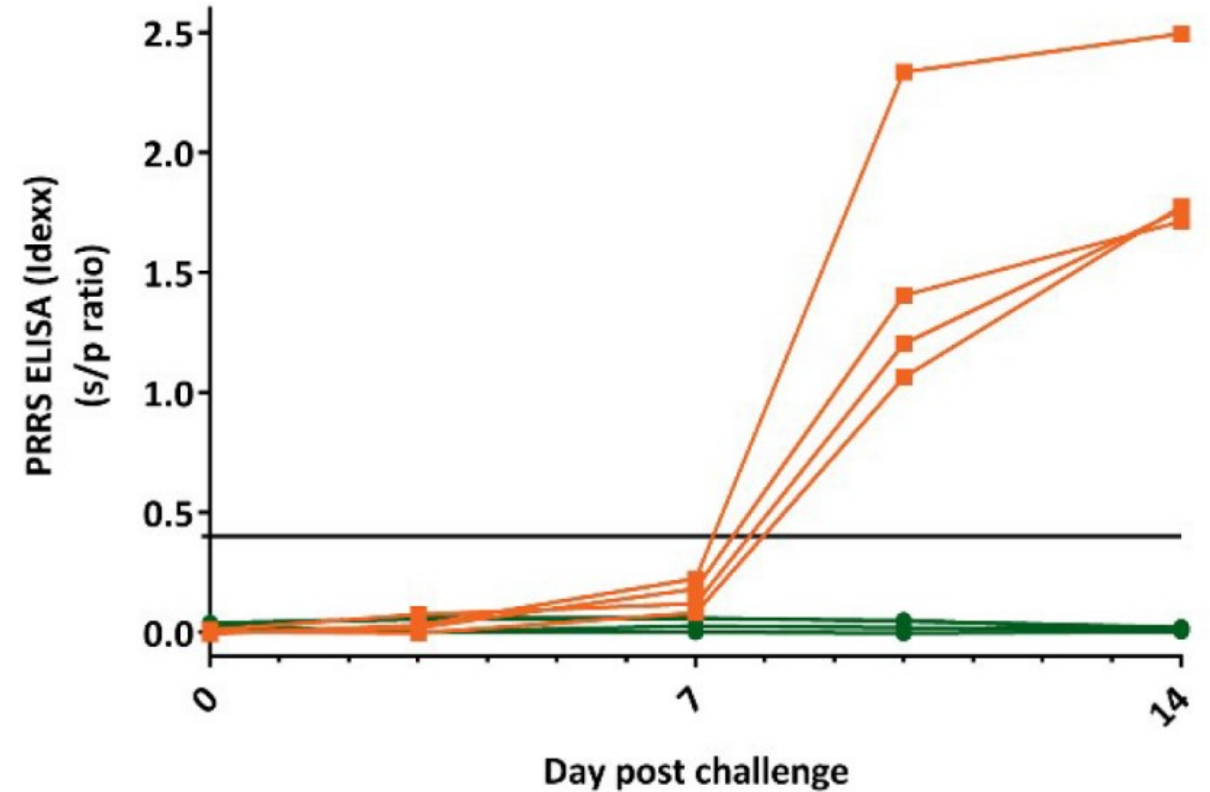




Consequences of viral disease

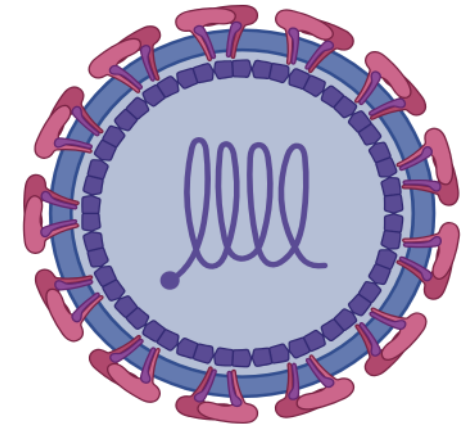
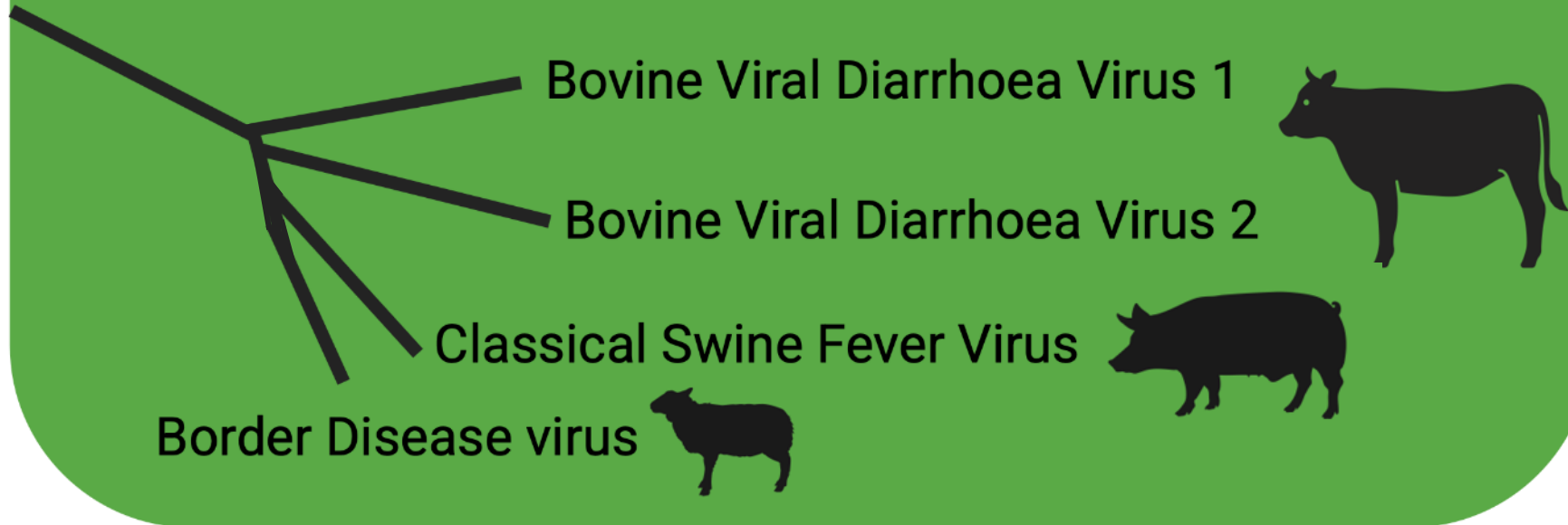


Porcine Reproductive and Respiratory Syndrome



Pestiviruses infect livestock

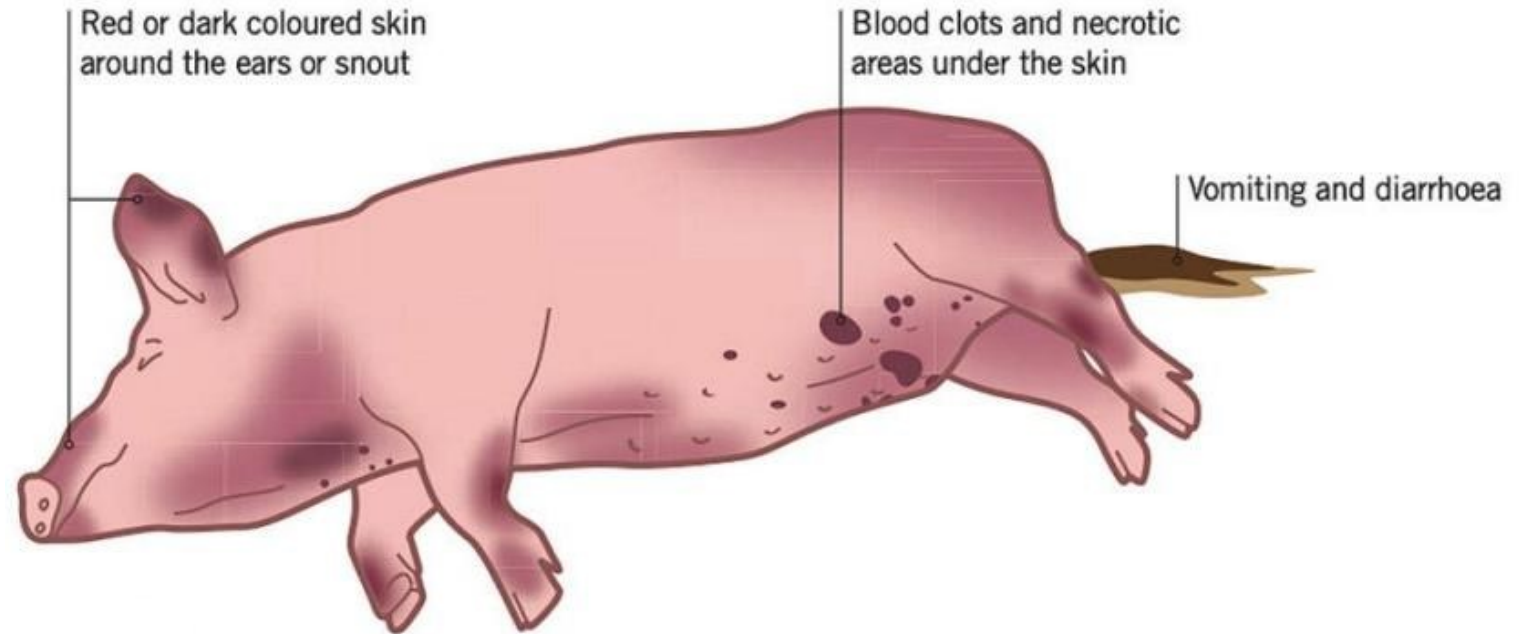
Pestiviruses



Classical swine fever (also known as Hog Cholera)

NOT to be confused with African Swine Fever or swine influenza

- High Fever
- Decreased Appetite
- Weakness, Incoordination, Paddling
- Red, blotchy skin
- Red ears
- Nasal and Eye Discharge
- Diarrhoea and Vomiting
- Coughing and difficulty Breathing
- Miscarriage, stillbirths, weak litters
- Congenital Tremor
- Death, which may be sudden

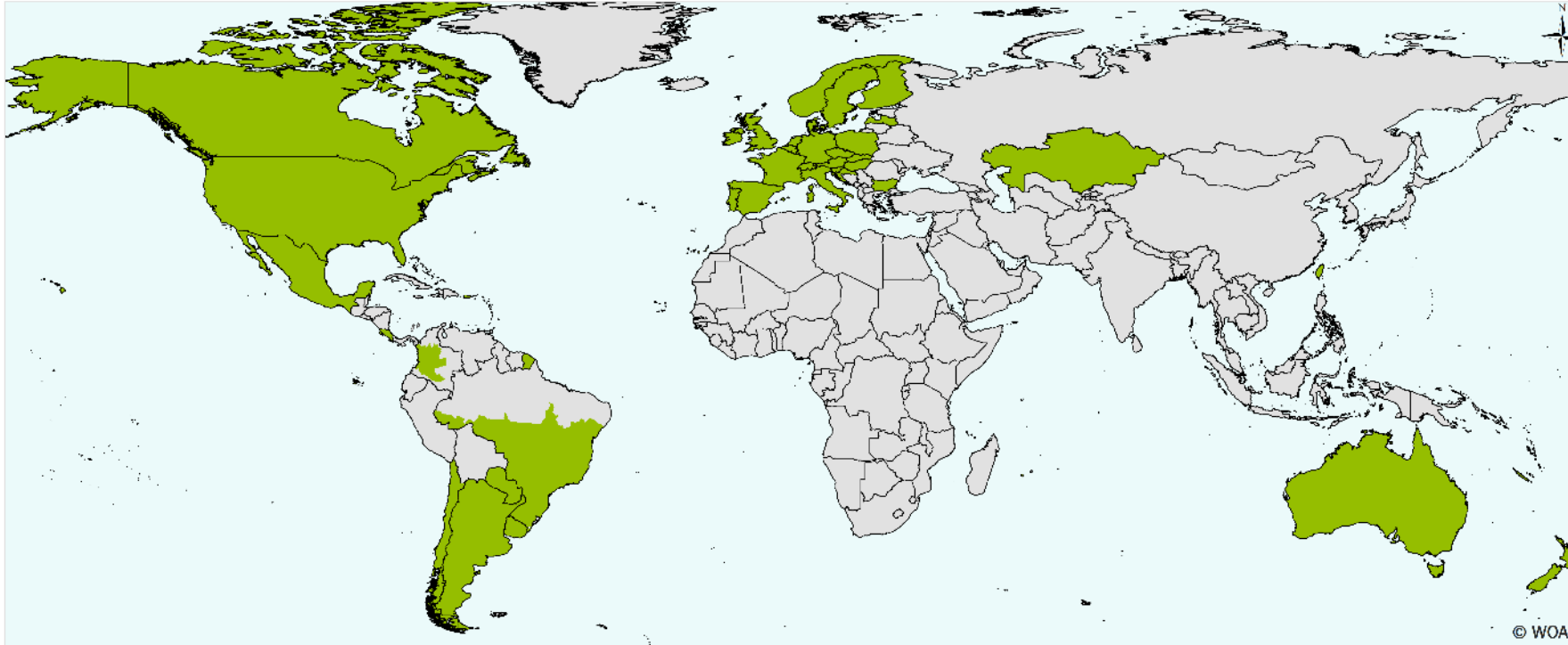




Big impact on animal welfare and productivity

Adapted from: Asian Agribiz Team, John Carr

Classical swine fever – a disease of global importance

Last update June 2025



-  Members and zones recognised as free from CSF
-  Countries and zones without an official status for CSF

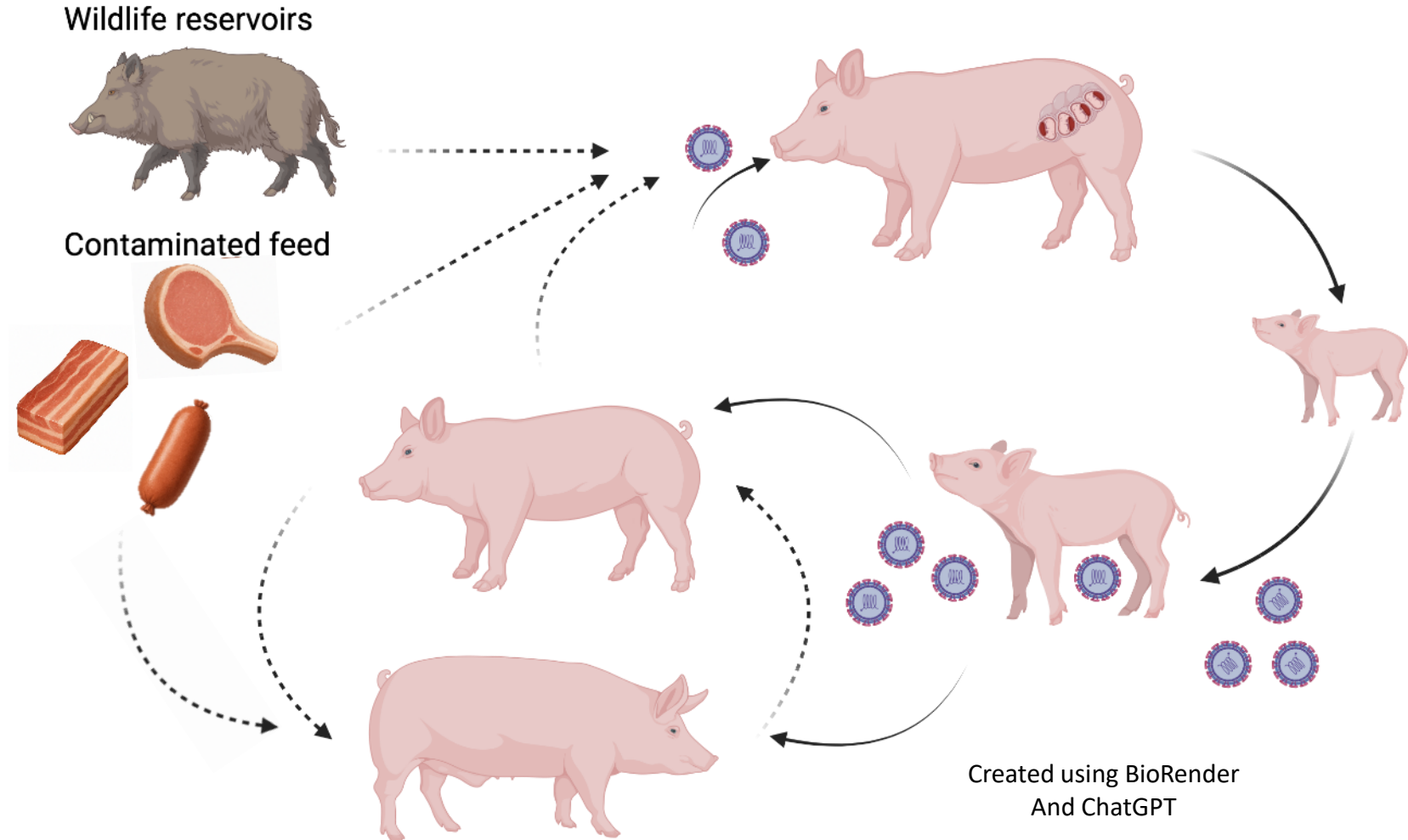
Eradicated in UK in 1966

**Outbreaks in: 1971
1986
2000**

**2000 UK outbreak
required cull of 75,000
pigs to contain it**

Source: World Organisation for Animal Health
(previously OIE)

Classical swine fever - transmission



Current management

- Vaccination
- Biosecurity
- Depopulation
- Trade restrictions

Pestiviruses interact with many host proteins



Review

Current Status of Genetically Modified Pigs That Are Resistant to Virus Infection

Hongming Yuan [†], Lin Yang [†], Yuanzhu Zhang, Wenyu Xiao, Ziru Wang, Xiaochun Tang, Hongsheng Ouyang ^{*} and Daxin Pang ^{*}
Viruses **2022**, *14*, 417. <https://doi.org/10.3390/v14020417>

CD46
Low density lipoprotein receptor
Heparan sulfate
ADAM17

Table 1. Host factors that act in CSFV infection.

Host Factors	Function in CSFV Infection	Promote (+) or Inhibit (−) CSFV Growth in Host Cell
Annexin2	Interacts with CSFV E2 and NS5A, promote CSFV replication	+
IFITM1-3 (interferon-induced trans membrane protein 3)	Modifies the membrane structure or alter endosomal physiology to impair viral membrane fusion	—
ARFGAP1 (ADP-ribosylation factor GTPase-activating protein 1)	Binds to CSFV NS5A and promote CSFV replication	+
β-actin	The amino acids 95-188 of β-actin are responsible for the interaction between β-actin and CSFV E2	+
Caveolin-1	CAV1-mediated endocytosis is necessary for CSFV invasion	+
NDP52 (nuclear dot protein 52)	CSFV inhibits NDP52 expression. Additionally, inhibiting NDP52 promotes interferon and TNF release, acting on the NF-κB pathway	+
GBP1 (guanylate-binding protein 1)	The N-terminal globular GTPase domain of GBP1 interacts with CSFV NS5A. Overexpression of GBP1 inhibits CSFV replication; knocking down GBP1 significantly promotes CSFV replication. Furthermore, the K51 of GBP1 is essential for CSFV replication	—
PSMB10 (proteasome subunit beta 10)	Acts as an NS3-interacting partner in CSFV infection. Overexpression of PSMB10 inhibited CSFV replication	+
POASL (interferon-inducible oligoadenylate synthetase-like protein)	Interacts with MDA5 to enhance MDA5-mediated type I IFN signaling and suppress CSFV replication	—
MERTK (Mer tyrosine kinase)	Interacts with CSFV E2 to facilitate CSFV entry, and down-regulates the expression of IFN-β to enhance CSFV replication	+
MG132	Activates JAK-STAT pathway and up-regulates several interferon-stimulated genes' (ISGs) expression in CSFV infection cells	—
RACK1 (receptor for activated C kinase 1)	RACK1 interacts with NS5A, inhibiting CSFV replication by inhibiting NF-κB activation	—
PRNF114 (porcine RING finger protein 114)	Interacts with NS4B and degrades NS4B through a proteasome-dependent pathway	—
Rab1b, Rab5, Rab7, and Rab11	Regulates CSFV endocytosis	+
Rab18	Interacts with NS5A and mediates virus replication and assembly	+
DCNT6 (dynactin subunit 6)	Interacts with E2, and the DCNT6-E2 interaction is important for CSFV replication and viral virulence	+
Torsin-1A	Interacts with E2, disrupting Torsin-1A-E2 interaction to completely inhibit CSFV replication	+
CCDC115 (coiled-coil domain-containing 115)	CCDC115-E2 interaction is essential for CSFV replication in swine macrophages	+
LamR (laminin receptor)	Acts as an alternative attachment receptor, interacting with E ^{ms}	+
Fatty acid synthase (FASN)	FASN participates in the formation of the replication complex. Knocking down FASN in host cells inhibits CSFV replication	+
PCBP1 (poly C-binding protein 1)	Interacts with N ^{pro} , down-regulating type I interferon in CSFV infection cells	+



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Journal of
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REPLICATION

1 October 2004 Volume 78 Issue 19
<https://doi.org/10.1128/jvi.78.19.10765-10775.2004>

Temporal Modulation of an Autoprotease Is Crucial for Replication and Pathogenicity of an RNA Virus

T. Lackner¹, A. Müller¹, A. Pankraz¹, P. Becher¹, H.-J. Thiel¹, A. E. Gorbalenya^{2,*}, N. Tautz^{1,*}



Dissection of a viral autoprotease elucidates a function of a cellular chaperone in proteolysis

T. Lackner, H.-J. Thiel, and N. Tautz[†]

Institut für Virologie (Fachbereich Veterinärmedizin), Justus-Liebig-Universität Giessen, Frankfurter Strasse 107, D-35392 Giessen, Germany

Edited by Charles M. Rice, The Rockefeller University, New York, NY, and approved November 18, 2005 (received for review September 21, 2005)



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Journal of
Virology®

GENOME REPLICATION AND REGULATION OF VIRAL GENE EXPRESSION

March 2019 Volume 93 Issue 5 10.1128/jvi.01714-18

<https://doi.org/10.1128/jvi.01714-18>

CRISPR/Cas9-Mediated Knockout of DNAJC14 Verifies This Chaperone as a Pivotal Host Factor for RNA Replication of Pestiviruses

O. Isken^a, A. Postel^b, B. Bruhn^a, E. Lattwein^{a,*}, P. Becher^b, N. Tautz ^a



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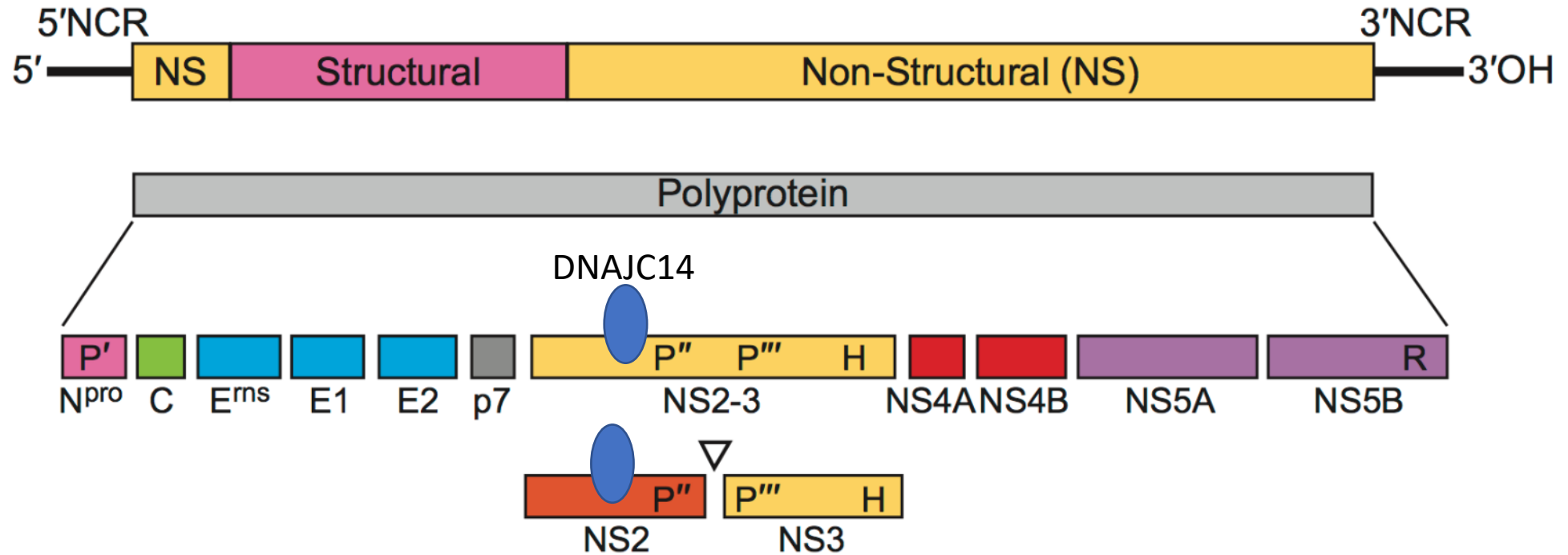


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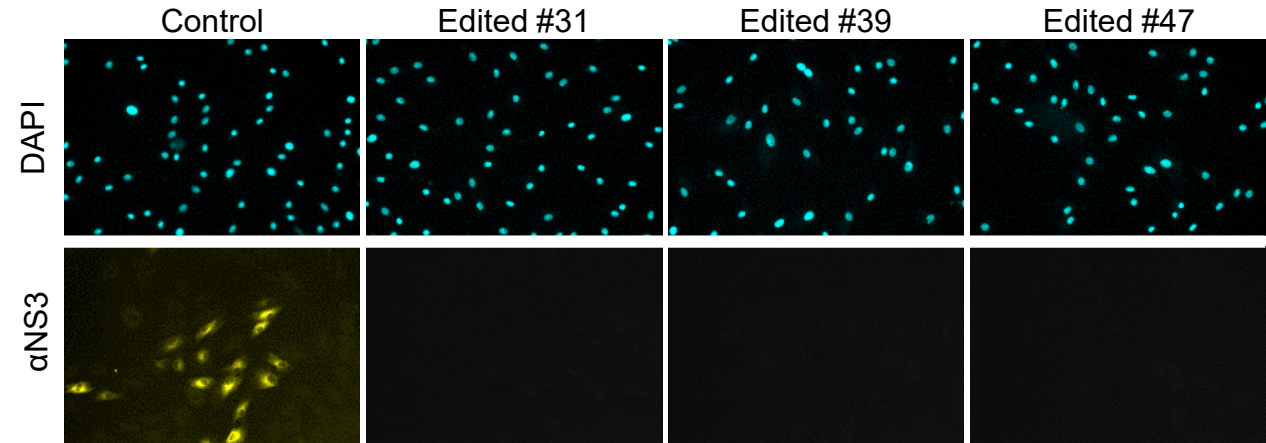
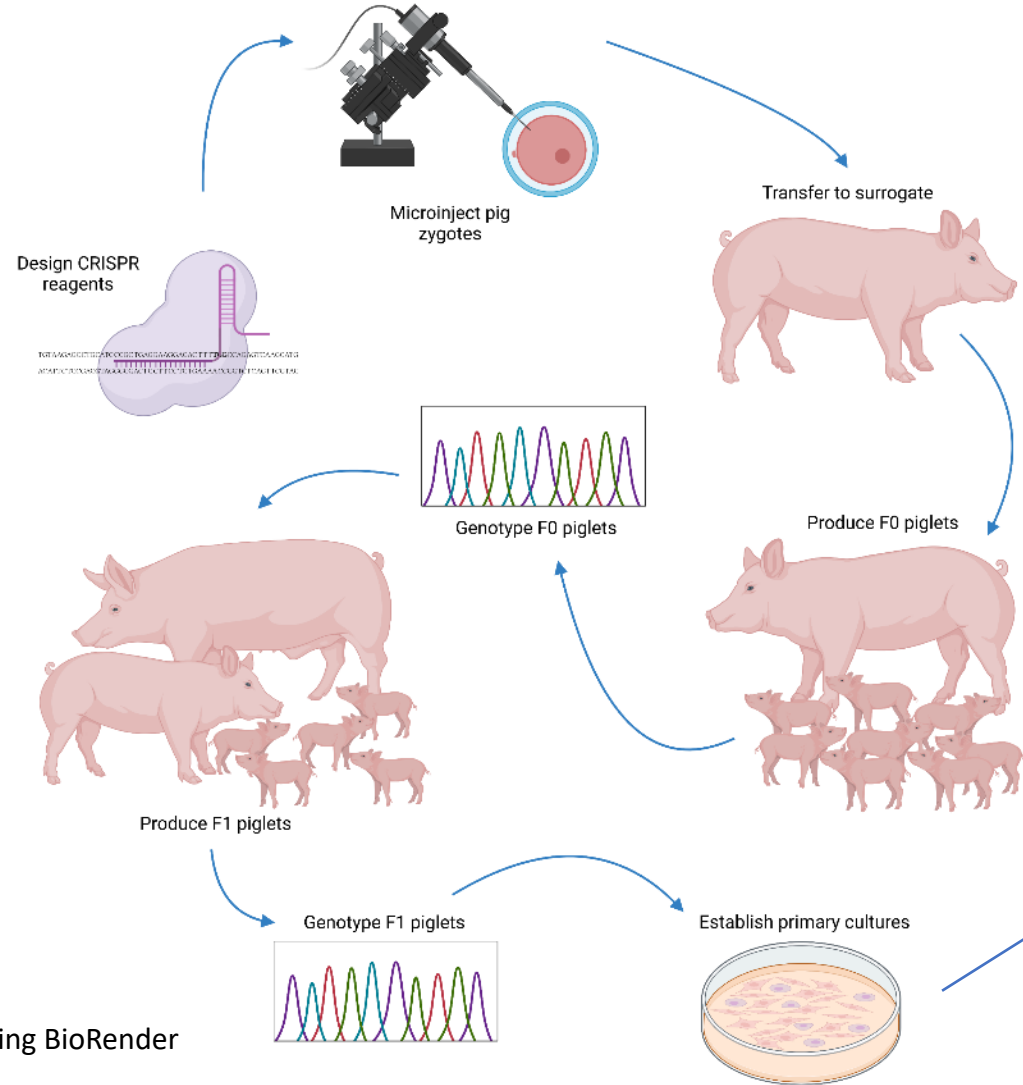
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Classical pestiviruses have a compact RNA genome



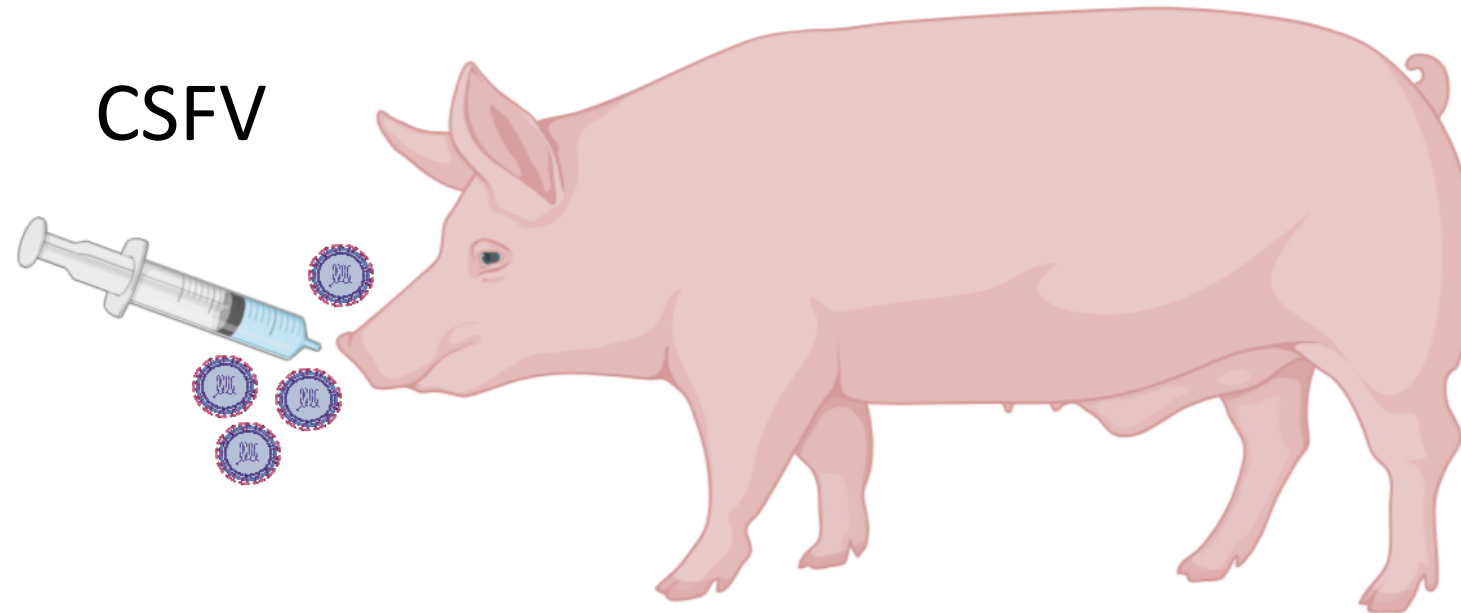
We produced pigs with edited DNAJC14

CSFV challenge



Created using BioRender

Is DNAJC14 essential for viral replication *in vivo*?



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4 x edited 4 x control

Monitored for 22 days

Clinical signs

Leukopenia

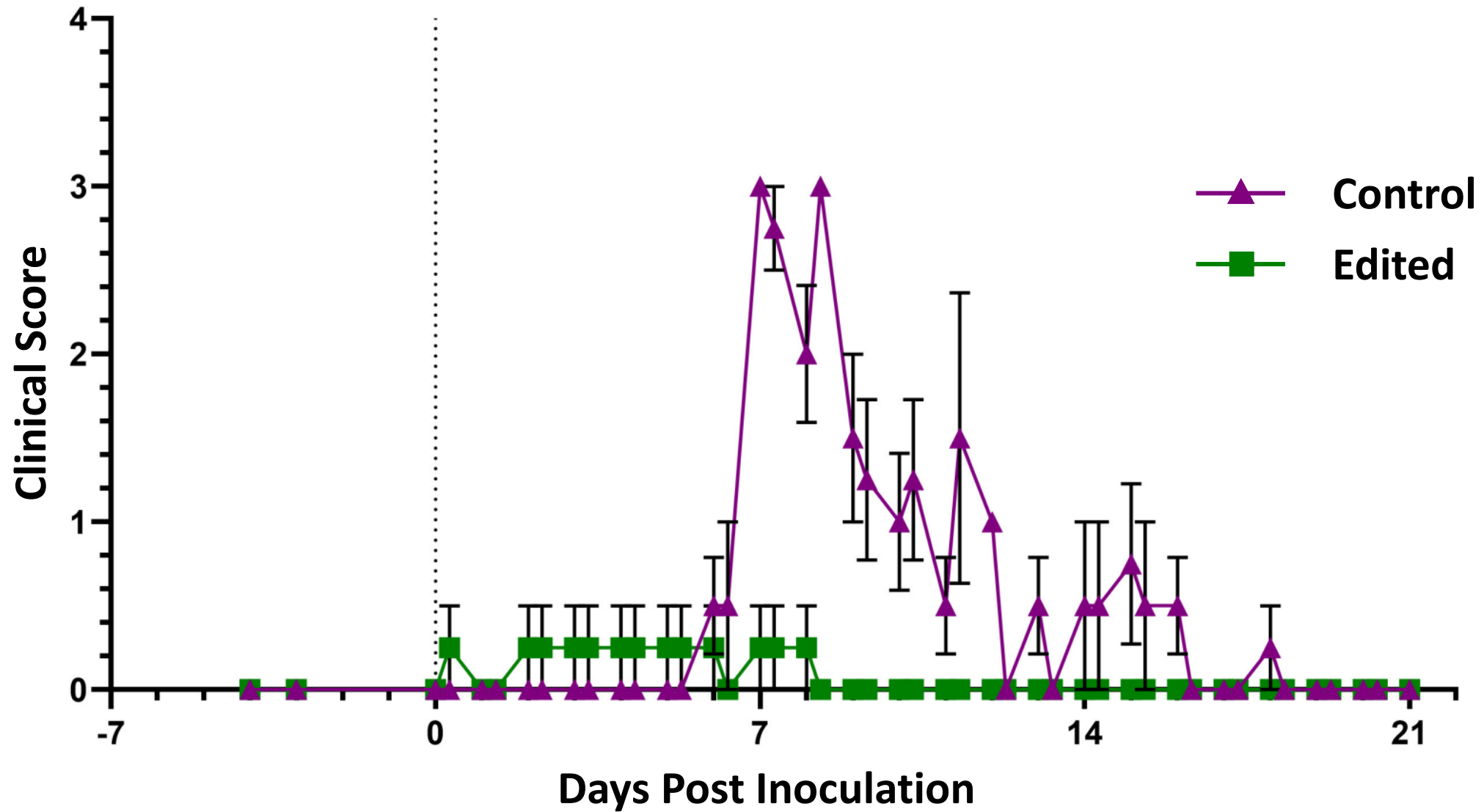
Viraemia

Antibodies

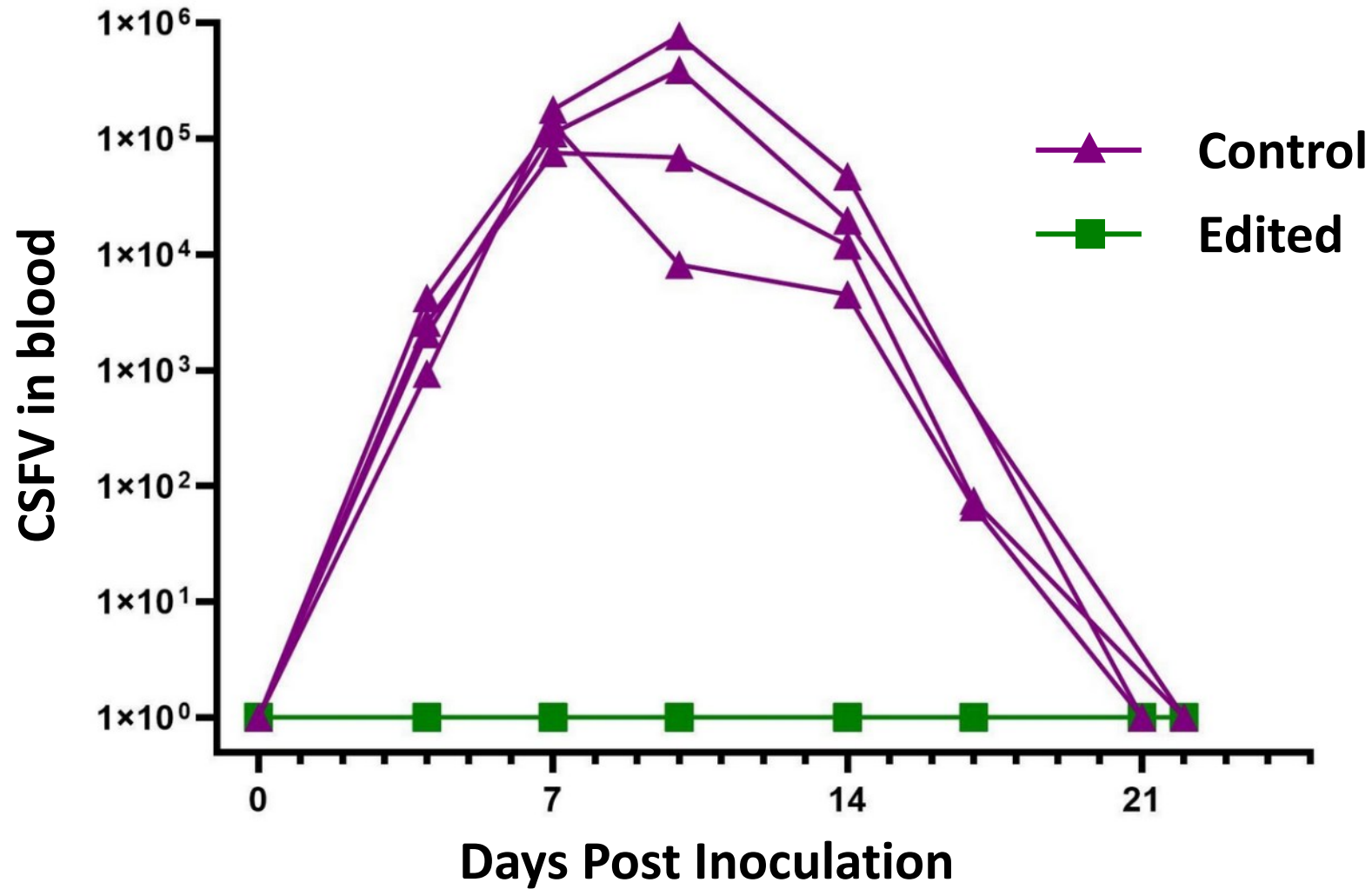
Shedding

Tissue PCR

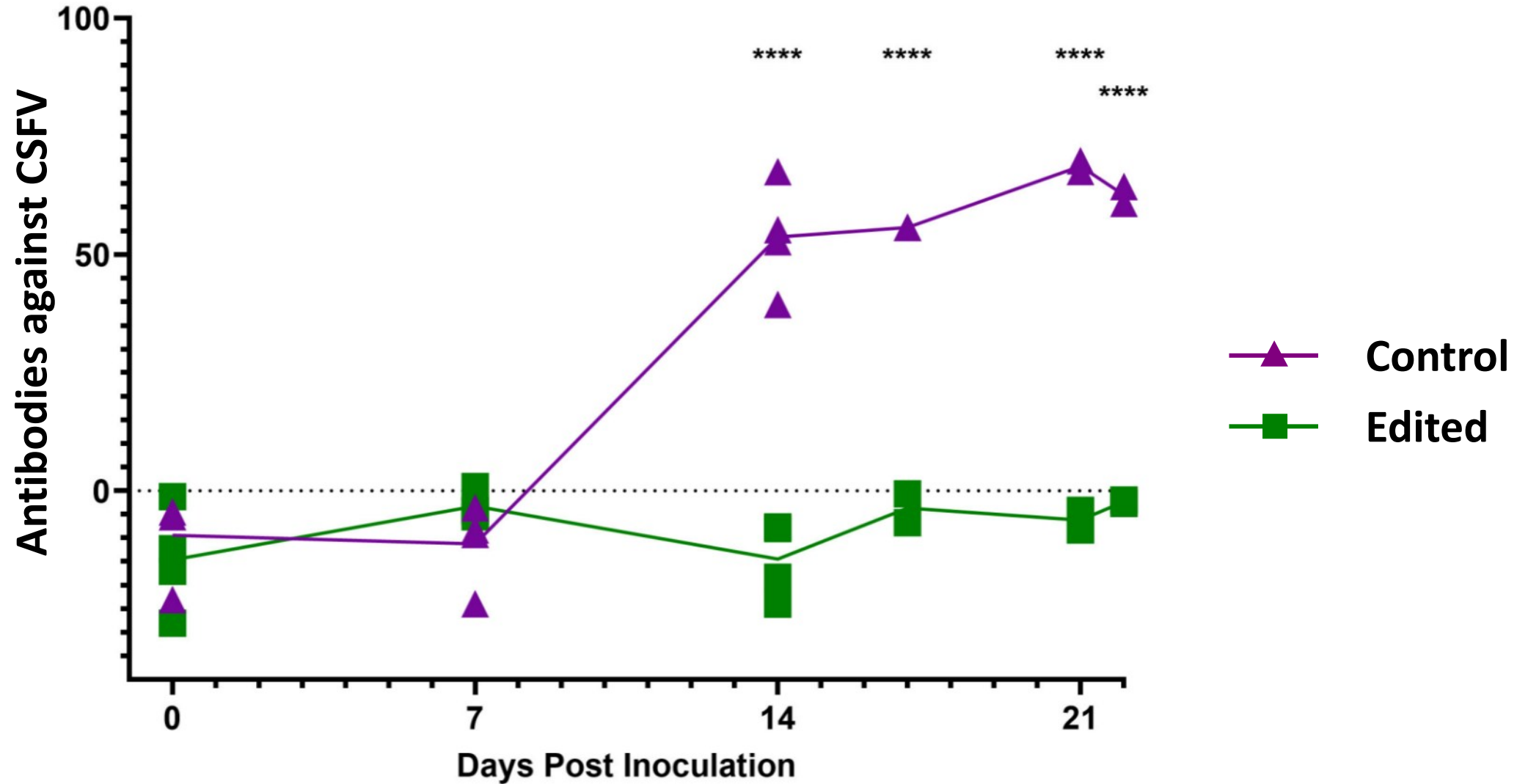
Edited pigs do not develop clinical signs



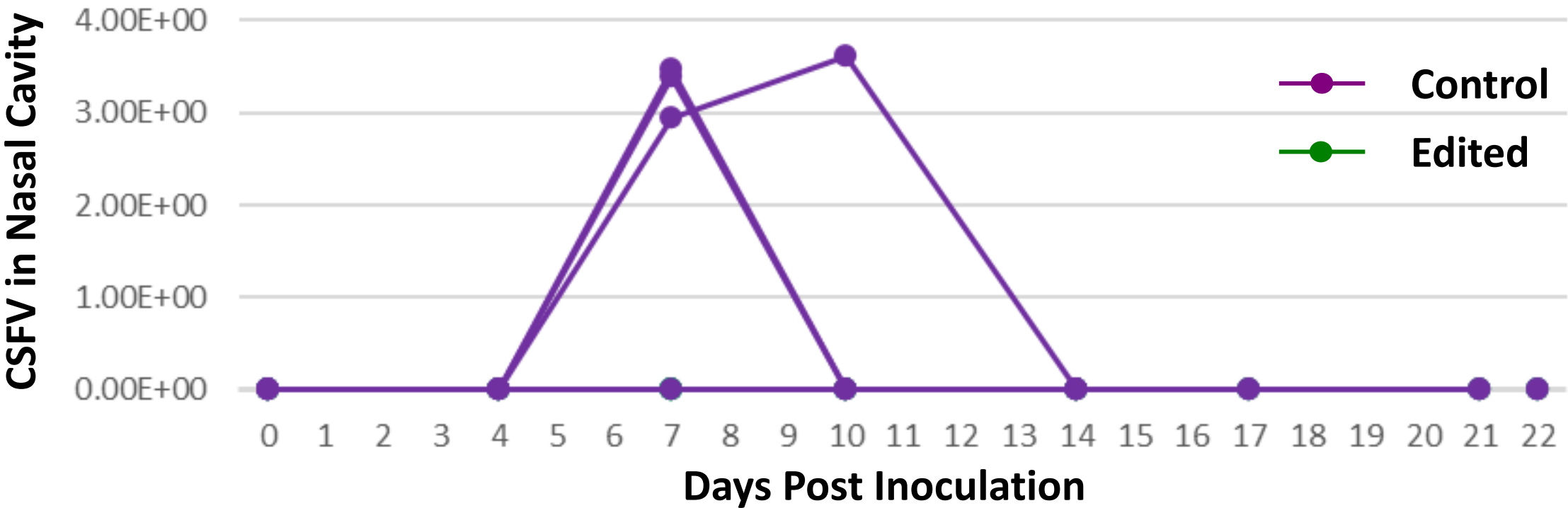
Edited pigs have no CSFV in their blood



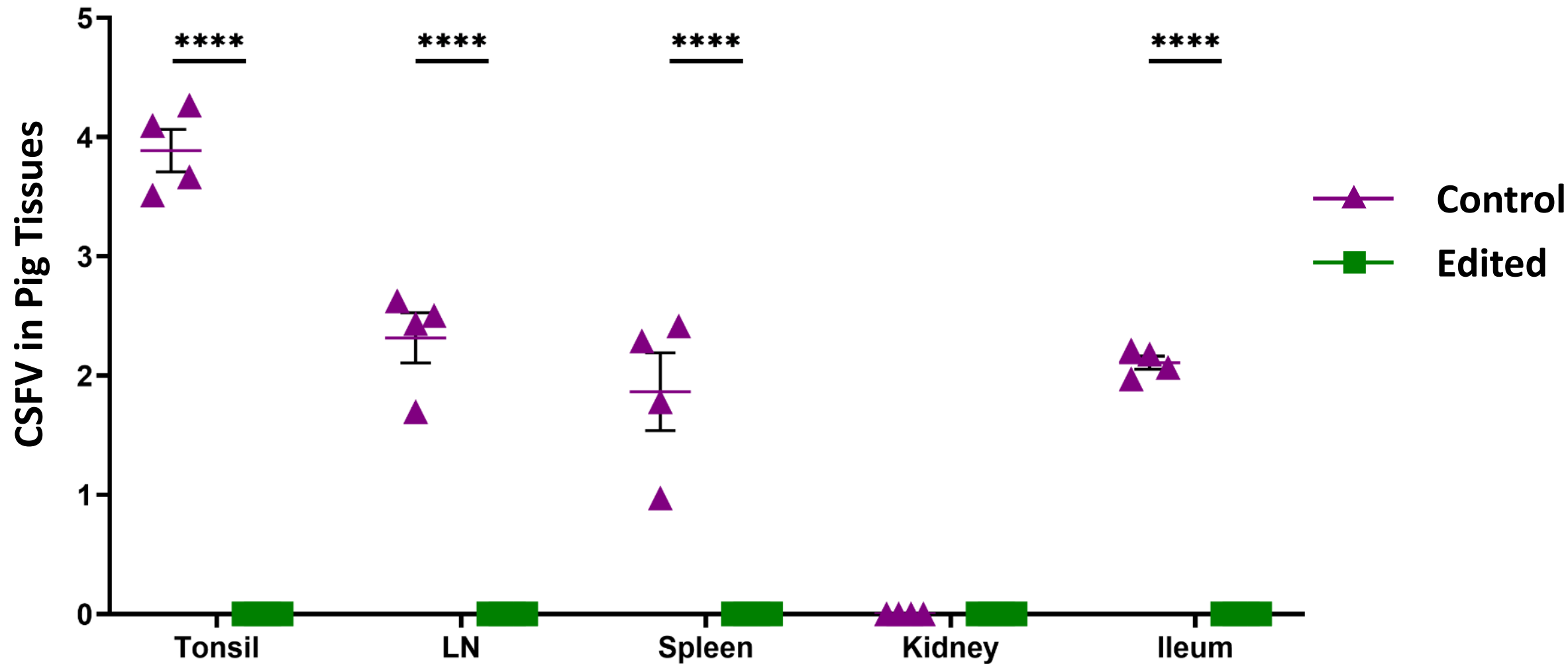
Edited pigs do not raise antibodies



Edited pigs do not shed CSFV



Edited pigs do not have CSFV in their tissues



Report

DNAJC14 gene-edited pigs are resistant to classical pestiviruses

Helen Crooke ^{1,4}, Stefanie Schwindt^{2,4}, Sarah L. Fletcher ³, Olaf Isken ², Sophie Harding ¹,
Nicholas Berkley ¹, Christine Tait-Burkard ³, Claire Warren³, C. Bruce A. Whitelaw ³,
Norbert Tautz ², and Simon G. Lillico ^{3,*}

doi: 10.1016/j.tibtech.2025.09.008.



- Application of NGTs has significant potential to contribute to the control of animal diseases
- Implementation could:
 - Improve welfare
 - Reduce reliance on antimicrobials
 - Reduce environmental footprint of animal agriculture
 - Support reliable production outcomes
- Failure to implement such technologies could be viewed as a biosafety failure

Thank you